

AMYLOID PROTEIN PROCESSING AND POTENTIAL TARGET OF ALZHEIMER DISEASE

Dr. Ranjana Dhar

Associate Professor, Deptt. of Physiology, Silchar Medical College, Silchar, Assam

Abstract

Human stem cell models serve as the potential to provide platforms for phenotypic screens to recognize candidate treatments and associated cellular pathways involved in the pathogenesis of Alzheimer disease (AD). Amyloid precursor protein (APP) processing and their derivatives like amyloid- β ($A\beta$) peptides generation are crucial processes involved in Alzheimer's disease (AD). Identification of modulators of APP processing was processed phenotypically to screen out a small-molecule in TS21 derived neurons. Avermectins identified as modulators in neural cells, can raise the relative production of non-toxic short $A\beta$ peptides in human cortical neurons at the expense of potentially more toxic longer peptides. Further, it was demonstrated that this avermectins effect is not only just because of interaction with the core γ -secretase and it is crucial for $A\beta$ production. Although this study reflects the avermectins could be prime targets of GABAA or glycine receptors, and may be possible reasons which affect $A\beta$ peptides processing.

Keywords: avermectin, amyloid, γ -secretase, neural cells, Alzheimer, pathogenesis

1. Introduction

Aging is a complex phenomenon that has been widely accepted as a major risk factor for the development of neurodegenerative disease like Alzheimer. Majority of population around the world are severely affected by Alzheimer disease (AD) (Bos et al. 2018), thereby; impact of this disease on public health system is deliberating growing. Alzheimer's disease cases were almost double every five years after age 65 (Corrada et al. 2008). The basic reason behind this critical situation is due to the lack of approved therapeutic measures as it concerning molecular mechanisms and actual cause are not fully understood (Ballard et al. 2011). Almost 70% cases of AD is actually caused by genetic level affecting various kinds of neuronal cell types in numerous genetic disorders conditions in human cells. For example; dopaminergic midbrain neurons in Parkinson's disease (Sanchez-Danes et al., 2012), and cortical neurons in Alzheimer's disease (AD) (Israel et al., 2012; Shi et al., 2012b; Yagi et al., 2011). Although AD prognosis can be studied with the help of cellular phenotypic character in appropriate human cell types, therefore it is now possible to understand the basic fundamental biology and its relevant disease-modifying pathways using genetic or pharmacological phenotypic screening in a relevant biological context. However, phenotypic screening of smaller compounds in cortical neurons which has potential to modify the production of amyloid- β ($A\beta$), an aggregation-prone and toxic peptide central cause of AD pathological conditions.

The aggregation of amyloid- β ($A\beta$) peptides results from proteolysis cleavage of amyloid precursor protein (APP); on account of rare autosomal dominant missense mutation in encoding

(APP) and presenilin (PSEN-1; PSEN-2) genes; which form catalytic subunits of γ -secretase complex (Ertekin-Taner, 2007). AD pathological condition arises because of *APP* gene over expression or chromosomal trisomy aberration at 21q position are responsible to cause uncontrolled production and accumulation of (A β) peptides (Rovelet-Lecrux et al. 2006; Sleegers et al. 2006,) and the early onset of Alzheimer disease pathology. In spite of these facts; it is striking note that no significant explanation was given which establish a concrete link in between the (A β) peptides aggregation and neuronal dysfunctions in AD pathology. Thereby; underlying amyloid hypothesis concern to amyloid processing and (A β) peptides production became a base of AD therapeutic approach in upcoming future studies. In this context, Findeis, (2007) and Kuperstein et al. (2010) showed the dynamic balance in longer and shorter (A β) peptides released after proteolytic cleavage by β -secretase and γ -secretase (Takami et al. 2009), as it is a more significant determining factor of AD disease progression and initiation compared to amyloid protein production. In reference to an effective strategy it would be necessary to regulate rather than inhibiting proteolytic processing of other substrates of γ -secretase. The possibility of APP processing augments in support of statement highlighting the association of carboxypeptidase activity of γ -secretase with its modulators; which could shift the A- β peptides production away from longer and more toxic species to shorter peptides, without affecting total A- β production or γ -secretase targeting of other substrates. While the results of larger clinical trials are yet to be concluded that modulators may be targeted one in AD patients (Soares et al. 2016; Toyn et al. 2016; Yu et al. 2014) and remain a promising possibility for further development. APP first undergoes processing through β and γ -secretase, and subsequently follow proteolysis step in a numerous cellular compartments (Small and Gandy, 2006). This complex events suggested that it may be possibly one of the reasons to modify amyloidogenic APP processing in a secretase-independent manner; which could alter longer A- β toxic peptide production toward shorter peptides. The present study goal is to detect secretase-independent, avermectins a smaller modulators of A- β processing that would elevate the production of A- β peptide fragments in human cortical neurons away from A β -42 to lesser toxic shorter peptides. These scientific data demonstrate that phenotypic screening in human stem cell models of AD; provides a potentially powerful strategy for identifying error prone disease-modifying pathways and associated compounds, independent of known approaches to modulate APP processing.

2. Materials and Methods

2.1 Experimental procedure

Preliminary screening and systematic studies were executed on neurons segregated from iPSCs isolated from those individuals with TS21 (Park et al. 2008). Additional other included genotypes were undertaken as non-diseased controls (NDCs; Israel et al. 2012), APP duplication (APPdup) (Israel et al. 2012), APP V717I (Moore et al. 2015) and PSEN1 M146I (Moore et al. 2015). iPSCs originator cells were cultured and retained as feeder free in essential-8 without supplying any antibiotics. iPSCs cell lines differentiation to cortical neurons was performed by assembling neural stem cells and follows the protocols as per previously described by Shi et al. (2012a), (2012c).

Neural stem cells assembling is essential at this stage as it minimize variability in developing neuronal cells usually seen during experimental procedure. Neural stem or progenitor cells were undertaken for subsequent independent neural differentiations for 15-30 days so as to produce postmitotic cortical neurons and astrocytes parallel for each trial or drug treatment. Each neuronal differentiation was treated separately as a biological replicate. Drug screening was tested on cortical neurons in 96-well plates (Greiner and Ibidi), while other additional experiments were tested in 12-well plates.

2.2 Immunocytochemistry

Cultured cells were fixed in paraformaldehyde 4% (w/v) in PBS and blocked with 5% normal donkey serum in 0.3% (v/v) Tween-20 in Tris-buffered saline before proceeding for immunofluorescent staining. Additionally, primary antibodies used in experimental procedure were anti-MAP2 (ab5392, Abcam), anti-CTIP2 (ab18465, Abcam), and anti-TBR1 (ab31940, Abcam), and one secondary antibody named as Alexa Fluor conjugated is used. Resultant stained cells were observed in an inverted confocal microscope (Olympus FV1000) and obtained results are imported for visualization purpose (PerkinElmer Velocity).

2.3 Drugs

Additional drugs such as (R)-flurbiprofen (Cayman), the γ -secretase modulator E2012 (Chem Express), abamectin (Santa Cruz Biotechnology), ivermectin (Tocris Biosciences), emamectin benzoate (Abcam), selamectin (MicroSource Discovery Systems), moxidectin (Santa Cruz), strychnine HCl (Abcam), picrotoxin (Tocris), muscimol (Tocris), and the γ -secretase inhibitors DAPT (Sigma) and L-685,458 (Merck-Millipore) are also included in this study. The Prestwick Chemical library supplied compounds at a concentration of 10mM in DMSO in a 96-well plates.

2.4 Drug Treatment

In preliminary screening, each drug was introduced at a final concentration of 1mM in 0.1% dimethyl sulpho-oxide (DMSO) in neural stem cell culture media. Additionally in other experimental procedure, drugs were mix-up in DMSO or water, and subsequently applied on neural cells in such a way, DMSO final concentrations did not surpass 0.4%. All drug treatments application started from 50 to 65 days after the beginning of neural cells induction, along with this, withdrawn media is further refreshed with the fresh media regularly at 48-hr intervals. Compound effects were normalized to taken as vehicle controls within each plate.

2.5 Biochemical Assays

Media supernatant prepared for biochemical analysis, it was centrifuge at 8000g to eliminate unwanted cellular debris as pellet, and supernatant stored at 20°C refrigerated conditions for further use. Other than this, measurement of Ab38, Ab40, and Ab42 was performed with the help of immunoassay (Meso Scale Diagnostics), and determined LDH activity to check the cell cytotoxicity activity with the help of (Roche) manufactured detection kit.

2.6 Immuno-precipitation

The A β peptide profile was determined by immunoprecipitation, using Ab-specific antibodies coupled to magnetic beads. Collected samples were studied as per previously designed by Portelius et al. (2007). Of the 25 identified A- β species, A- β 1-19 was found to be highly variable samples within the same treatment groups therefore, A- β 1-19 were not included for further analysis.

2.7 Immunoblotting detection

After collection in ice-cold PBS buffer solution, cells were further lysed in RIPA buffer containing 1 mM DTT, protease and phosphatase inhibitors (Pierce), and 25 U/mL DNase. The cell lysates soluble fraction was further subjected to SDS-PAGE and immunoblotting techniques. N-cadherin (610921, BD Transduction Laboratories), APP (SIG-39152, Covance), and histone H3 (ab1791, Abcam) were used as primary antibodies for detection purposes.

2.8 Cell-free γ -Secretase activity assay

The γ -secretase *in vitro* activity assay was analyzed as per Szaruga et al. (2015) described procedure.

2.9 Statistical Analysis

Statistical analysis was done with help of IBM SPSS and GraphPad Prism softwares. Analysis was performed to set to 0.05 level for all significant observation.

3. Results and Discussion

3.1 A Preliminary Phenotypic Screening to detect modifiers of A- β production

In this study phenotypic screening of neural stem cells originated from cultured TS21, (iPSC) induced pluripotent stem cells in 96-microwell plates to detect the presence of small molecule of modifiers of A- β peptides in cortical neuron of humans, (fig.1). Genetically modified neuronal cells can have ability to over produce A- β type peptides by Shi et al. (2012b), and thus they are sensitive towards drug screening purpose. A single point screening of amyloid peptides (A- β -38, A- β -40, and A- β -42) was done at 1mM concentration of drugs treatment after 4 days immunoassay was performed to check these antibodies presence and in between these events, used media was exchanged regularly with fresh media after 48h time interval. Thereafter activity of lactate dehydrogenase (LDH) in extracellular media was determined after 6 days of drugs treatment as an indicator of cellular toxicity. Each microtitre well plate performance was determined as per coefficient of variation (CoV) of DMSO as control (n=5 culture per plate). In this context, obtained mean (CoV) value for A- β 38/A- β 42 (9.11%), and A- β 40/A- β 42 (7.579%) respectively. These mean values reflects the minimal level of variations in A- β ratios in control sample thereby, it is used to quantify the targeted compounds. A control-independent method was used to detect target compounds with the help of given B-score in order to find out the positional differences within the 96-well plates (Brideau et al. 2003). Adjustment in B-Score can be deduced by using the open-source Bioconductor cell HTS2 package (Boutros et al. 2006).

The present study results shows the LDH activity was elevated in seventy three compounds as it has higher B-score (B score >3) and similar score results were obtained in case of 55 compounds as they reflects the reduction in a A β 42 production relative to total A β production. Similar kind of validation of further compounds identification was repeated again at 1mM in triplicate manner, and significant Fisher's least significant difference test was performed to reproduce their initial effect or at least two of three replicates reflect increment in >10% in the A β 38/A β 42 or A β 40/A β 42 ratio compared to DMSO treatment. Additionally among all tested compounds only two compounds were shows the validated hits as these hits (0.1666%) reflects increment in A β 38 to A β 42 ratio in a dose-dependent manner.

3.2 Avermectins alter the proportion of A β 38/A β 42 ratio in humancortical neurons

Avermectins members class compounds with non-steroidal anti-inflammatory drugs (NSAIDs) category GSM (R)-flurbirofen and the GSM E-2012 can have ability to raise the A β 38 to A β 42 ratio in TS21 cortical neurons in a dose-dependent manner ($F(7, 16) = 43.766, p < 0.0001$; $F(7, 16) = 1633$ at $p < 0.0001$, respectively). Other than this one of the identified compound named as abamectin had minimal level of A β 38/A β 42 increment ($F(5, 15) = 2.754, p = 0.0586$), whereas, structural analog compounds such as ivermectin significantly raised the A β 38/A β 42 level in a dose dependent manner ($F(5, 15) = 4.435, p = 0.0221$) respectively. Additionally, avermectin related family compounds namely Emamectin benzoate and selamectin had shown similar effect on A β 38/A β 42 level, as per obtained results of these identified compounds; Emamectin benzoate had shown a moderate effect ($F(5, 15) = 6.284, p = 0.0050$) while selamectin shown good potency towards the A β 38/A β 42 ratio ($F(5, 15) = 16.18, p = 0.0003$).

All other avermectin related compounds such as moxidectin, a member of macrocyclic lactones do not show any significant effect on A β 38/A β 42 ratio as reflected through ($H(5) = 11.77, p = 0.0380$; however, for all post hoc comparisons is also made at $p > 0.05$).

All the recorded results of the study had shown that most of the avermectin belonging family of compounds can have higher A β 38/A β 42 ratio by increasing A β 38 and/or reducing A β 42, while few of them had shown additional effects on A β 40 peptides.

Abamectin, ivermectin, and selamectin had significantly raise A β 38 level ($F(5, 15) = 6.543, p = 0.0040$; $F(5, 15) = 13.05, p = 0.0003$; $H(5) = 16.59, p = 0.0053$, respectively) comparative to A β 40 whereas; Emamectin benzoate and selamectin significantly decreased A β 42 level ($F(5, 15) = 5.889, p = 0.0033$; $F(5, 15) = 7.22, p = 0.0039$, respectively). In addition, abamectin, ivermectin, and selamectin had shown similar decrement in A β 40 level ($F(5, 15) = 6.063, p = 0.0058$; $H(5) = 12.35, p = 0.0303$; $F(5, 15) = 53.38, p = 0.0003$, respectively). However, all macrocyclic lactones had a greater tendency of lipophilicity thereby; it was concluded that the avermectins family related compounds had significant effect on A β production and it was noteworthy this effect is on account of membrane stress and or due to γ -secretase-APP interaction. Previously encoded study results of (Prichard et al. 2012) reflect that it had no significant correlation between the studied compounds efficacy and lipophilicity on the basis of clogp values. From this results it was concluded that avermectin, selamectins shows higher potent nature as

per clogp values (6.0 and 6.3) respectively while other two compounds such as milbemycin and moxidectin had least values <3.0 clogp value

While avermectins abamectin and ivermectin had a moderately relative values as per recorded cLogP values of 5.3 and 4.8, respectively (Prichard et al. 2012). As per these observed results it was noteworthy that the avermectin effect on A β production is not only just because of their own lipophilicity nature, but it requires additional observations to detect possible molecular targets.

3.3 Effects of Avermectin on APP Processing and A β Peptide Production

As per the obtained fraction of A β peptides produced from human cortical neurons after A β 38/A β 42 immunoassay. The studied 3 A β peptides fraction originated on account of proteolysis of APP stimulated with the effect of avermectin compounds and A β peptides fraction were further analyzed with the help of immunoprecipitation. Previously encoded studies had consistent increment in A β peptides such as A β 1-14, A β 1-15, and A β 1-16, while A β 1-34 peptide shown decrement when APP processed with a γ -secretase inhibitor (GSI) (Portelius et al. 2010a, 2012) while GSM modifiers treatment leads to an increment in A β 1-37 peptide along with decrement in A β 1-39, A β 1-40, and A β 42 peptides (Portelius et al. 2010b, 2014). As per immunoassay and earlier studied results A β 1-14, A β 1-15, A β 1-16, A β 1-34, A β 1-37, A β 1-38, A β 1-39, and A β 1-40 peptides were quantified for further analysis except A β 1-42 as it was not reliably detected and quantifiable in all the undertaken samples. Although selamectin (1.5 mM) treatment to TS21 neurons over ten days had significantly raised the A β 1-37 and a decrease in A β 1-40 level, but it has least effects on A β 1-14, A β 1-15, and A β 1-16. This increment and decrement in the level of A β 1-37 and A β 1-40 is occurred consistently as with γ -secretase modulation (Portelius et al., 2010b), while it has shown minimal degree of effect on A β 1-14, A β 1-15, and A β 1-16 peptides as it may be due to significant of γ -secretase activity (Portelius et al. 2010a, 2012).

Immunoassay experiment reflect avermectin induced a significant effect on A β -38 comparative to the A β 40/42. It is just because of no significant effect of A β -38 (Portelius et al. 2010b) on application of GSM E2012 thereby; A β 38/A β 42 complex system is more sensitive towards immunoassay detection comparative to graded IP-MALDI technique, particularly for low-abundance and relatively hydrophobic peptides.

3.4 Effect of Avermectins on A β peptides production and its unusual effect on ion channels

This study reflects that avermectin has significant effect on high affinity glutamate-gated chloride channels. Inflow of Cl⁻ ions stabilize the open conformations of ion channels in neural cells adversely shown paralyzing effect on nematodes and other parasites (Wolstenholme and Rogers, 2005). In addition, avermectins ($F(2, 41) = 176.8, p < 0.0001$) also shown agonists effect on ligand-gated chloride channels *i.e.* γ -aminobutyric acid-A (GABA_A) and glycine at nanomolar concentration (Dawson et al. 2000) as similarly shown by and picrotoxin ($F(5, 41) = 9.996, p < 0.0001$). Other than this, the most potent GABA receptor agonist namely muscimol (100 mM) had no significant effect on the A β 38/A β 42 level ($F(6, 17) = 2.007, p = 0.1209$) Shan et al. (2001).

In this study GABAA and glycine receptor activity was tested on the basis of the A β 38/A β 42 level as this A β level is depicted through ivermectin (1 mM) and selamectin (1 mM) absence or presence. Two-way ANOVA analysis showed the significant main effects of the avermectins (F(2, 41) = 176.8, $p < 0.0001$) and picrotoxin (F(5, 41) = 9.996, $p < 0.0001$) on A- β peptides. Although it has no significant connection in between the avermectins and picrotoxin (F(10, 41) = 0.8645, $p = 0.5724$). Similarly another muscimol; an inducer of GABAA receptor was tested up to 100 mM; and it had no significant effect on the A β 38/A β 42 level (F(6, 17) = 2.007, $p = 0.1209$). In addition glycine receptor antagonist such as strychnine (0.3–30 mM) in the presence and absence of these avermectins tested at 1 Mm had significant effect (F(2, 41) = 43.55, $p < 0.0001$) but do not show any interaction between these two compounds (F(10, 41) = 0.6173, $p = 0.7901$). In addition, no significant inhibition of the concentration-dependent effect of ivermectin or selamectin on the A β 38/A β 42 level was observed when dose response was performed in the presence of strychnine. These study findings clearly indicate that avermectins could be prime targets of GABAA or glycine receptors, and may be affect A β peptides processing.

4. Conclusion

This study concluded that the human cortical neurons derived from an individual with induced pluripotent stem cell as TS21 highlighted the probability of identifying phenotypically screening of a small-nontoxic molecule derived from longer toxic form of A β peptides responsible to cause AD in humans. In this study, avermectins as modulators were identified a prime precursor of A β peptides production, which act independently of γ -secretase to alter APP processing in a manner similar to γ -secretase modulation. The recent emergence and continued development of human cell models of disease in combination with traditional phenotypic screening approaches open a new avenue to allow the identification of potential drug candidates in addition to uncovering new pathways concerning with Alzheimer disease pathology.

5. References

1. Bos, D., Wolters, F.J., Darweesh, S.K.L., Vernooij, M.W., de Wolf, F., Ikram, M.A., & Hofman, A. (2018). Cerebral small vessel disease and the risk of dementia: A systematic review and meta-analysis of population-based evidence. *The Journal of the Alzheimers Association*. 14(11):1482-1492.
2. Corrada, M.M., Brookmeyer R., Berlau D., Paganini-Hill A. & Hawas C.H. (2008). Prevalence of dementia after age 90. *Neurology*, 71 (5) 337-343.
3. Ballard, C.; Gauthier, S.; Corbett, A.; Brayne, C.; Aarsland, D.; Jones, E. Alzheimer's disease. *Lancet* 2011, 377, 1019–1031.
4. Sanchez-Danes, A., Richaud-Patin, Y., Carballo-Carbajal, I., Jimenez-Delgado, S., Caig, C., Mora, S., Di Guglielmo, C., Ezquerro, M., Patel, B., Giralt, A., et al. (2012). Disease-specific phenotypes in dopamine neurons from human iPS-based models of genetic and sporadic Parkinson's disease. *EMBO Mol. Med.* 4, 380-395.

5. Shi, Y., Kirwan, P., Smith, J., MacLean, G., Orkin, S.H., and Livesey, F.J. (2012). A human stem cell model of early Alzheimer's disease pathology in Down syndrome. *Sci. Transl. Med.* 4, 124ra129.
6. Israel, M.A., Yuan, S.H., Bardy, C., Reyna, S.M., Mu, Y., Herrera, C., Hefferan, M.P., Van Gorp, S., Nazor, K.L., Boscolo, F.S., et al. (2012). Probing sporadic and familial Alzheimer's disease using induced pluripotent stem cells. *Nature* 482, 216-220.
7. Ertekin-Taner, N. (2007). Genetics of Alzheimer's disease: a centennial review. *Neurol. Clin.* 25, 611–667.
8. Rovelet-Lecrux, A., Hannequin, D., Raux, G., Le Meur, N., Laquerriere, A., Vital, A., Dumanchin, C., Feuillette, S., Brice, A., Vercelletto, M., et al. (2006). APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. *Nat. Genet.* 38, 24–26.
9. Sleegers, K., Brouwers, N., Gijssels, I., Theuns, J., Goossens, D., Wauters, J., Del-Favero, J., Cruts, M., van Duijn, C.M., and Van Broeckhoven, C. (2006). APP duplication is sufficient to cause early onset Alzheimer's dementia with cerebral amyloid angiopathy. *Brain* 129, 2977-2983.
10. Findeis, M.A. (2007). The role of amyloid beta peptide 42 in Alzheimer's disease. *Pharmacol. Ther.* 116, 266–286.
11. Kuperstein, I., Broersen, K., Benilova, I., Rozenski, J., Jonckheere, W., Debulpaep, M., Vandersteen, A., Segers-Nolten, I., Van Der Werf, K., Subramaniam, V., et al. (2010). Neurotoxicity of Alzheimer's disease Aβ peptides is induced by small changes in the Aβ₄₂ to Aβ₄₀ ratio. *EMBO J.* 29, 3408-3420.
12. Takami, M., Nagashima, Y., Sano, Y., Ishihara, S., Morishima-Kawashima, M., Funamoto, S., and Ihara, Y. (2009). γ-Secretase: successive tripeptide and tetrapeptide release from the transmembrane domain of beta-carboxyl terminal fragment. *J. Neurosci.* 29, 13042-13052.
13. Boutros, M., Bras, L.P., and Huber, W. (2006). Analysis of cell-based RNAi screens. *Genome Biol.* 7, R66.
14. Brideau, C., Gunter, B., Pikounis, B., and Liaw, A. (2003). Improved statistical methods for hit selection in high-throughput screening. *J. Biomol. Screen.* 8, 634–647.
15. Dawson, G.R., Wafford, K.A., Smith, A., Marshall, G.R., Bayley, P.J., Schaeffer, J.M., Meinke, P.T., and McKernan, R.M. (2000). Anticonvulsant and adverse effects of avermectin analogs in mice are mediated through the gamma-aminobutyric acid (A) receptor. *J. Pharmacol. Exp. Ther.* 295, 1051-1060.
16. Israel, M.A., Yuan, S.H., Bardy, C., Reyna, S.M., Mu, Y., Herrera, C., Hefferan, M.P., Van Gorp, S., Nazor, K.L., Boscolo, F.S., et al. (2012). Probing sporadic and familial Alzheimer's disease using induced pluripotent stem cells. *Nature* 482, 216-220.
17. Moore, S., Evans, L.D., Andersson, T., Portelius, E., Smith, J., Dias, T.B., Saurat, N., McGlade, A., Kirwan, P., Blennow, K., et al. (2015). APP metabolism regulates tau proteostasis in human cerebral cortex neurons. *Cell Rep.* 11, 689–696.

18. Park, I.H., Arora, N., Huo, H., Maherali, N., Ahfeldt, T., Shimamura, A., Lensch, M.W., Cowan, C., Hochedlinger, K., and Daley, G.Q. (2008). Disease-specific induced pluripotent stem cells. *Cell* 134, 877–886.
19. Portelius, E., Tran, A.J., Andreasson, U., Persson, R., Brinkmalm, G., Zetterberg, H., Blennow, K., and Westman-Brinkmalm, A. (2007). Characterization of amyloid beta peptides in cerebrospinal fluid by an automated immunoprecipitation procedure followed by mass spectrometry. *J. Proteome Res.* 6, 4433–4439.
20. Portelius, E., Zetterberg, H., Dean, R.A., Marcil, A., Bourgeois, P., Nutu, M., Andreasson, U., Siemers, E., Mawuenyega, K.G., Sigurdson, W.C., et al. (2012). Amyloid-beta(1-15/16) as a marker for gamma-secretase inhibition in Alzheimer's disease. *J. Alzheimers Dis.* 31, 335–341.
21. Portelius, E., Van Broeck, B., Andreasson, U., Gustavsson, M.K., Mercken, M., Zetterberg, H., Borghys, H., and Blennow, K. (2010b). Acute effect on the A β isoform pattern in CSF in response to gamma-secretase modulator and inhibitor treatment in dogs. *J. Alzheimers Dis.* 21, 1005–1012.
22. Portelius, E., Appelkvist, P., Stromberg, K., and Högglund, K. (2014). Characterization of the effect of a novel gamma-secretase modulator on A β : a clinically translatable model. *Curr. Pharm. Des.* 20, 2484–2490.
23. Prichard, R., Menez, C., and Lespine, A. (2012). Moxidectin and the avermectins: consanguinity but not identity. *Int. J. Parasitol. Drugs Drug Resist.* 2, 134–153.
24. Portelius, E., Dean, R.A., Gustavsson, M.K., Andreasson, U., Zetterberg, H., Siemers, E., and Blennow, K. (2010a). A novel A β isoform pattern in CSF reflects gamma-secretase inhibition in Alzheimer disease. *Alzheimers Res. Ther.* 2, 7.
25. Szaruga, M., Veugelen, S., Benurwar, M., Lismont, S., Sepulveda-Falla, D., Lleo, A., Ryan, N.S., Lashley, T., Fox, N.C., Murayama, S., et al. (2015). Qualitative changes in human gamma-secretase underlie familial Alzheimer's disease. *J. Exp. Med.* 212, 2003–2013.
26. Soares, H.D., Gasior, M., Toyn, J.H., Wang, J.S., Hong, Q., Berisha, F., Furlong, M.T., Raybon, J., Lentz, K.A., Sweeney, F., et al. (2016). The gamma secretase modulator, BMS-932481, modulates A β peptides in the plasma and CSF of healthy volunteers. *J. Pharmacol. Exp. Ther.* 358, 138–150.
27. Small, S.A., and Gandy, S. (2006). Sorting through the cell biology of Alzheimer's disease: intracellular pathways to pathogenesis. *Neuron* 52, 15–31.
28. Shi, Y., Kirwan, P., and Livesey, F.J. (2012a). Directed differentiation of human pluripotent stem cells to cerebral cortex neurons and neural networks. *Nat. Protoc.* 7, 1836–1846.
29. Shi, Y., Kirwan, P., Smith, J., MacLean, G., Orkin, S.H., and Livesey, F.J. (2012b). A human stem cell model of early Alzheimer's disease pathology in Down syndrome. *Sci. Transl. Med.* 4, 124ra129.
30. Toyn, J.H., Boy, K.M., Raybon, J., Meredith, J.E., Roberston, A.S., Guss, V., Hoque, N., Sweeney, F., Zhuo, X., Clarke, W., et al. (2016). Robust translation of GSM pharmacology across preclinical species and human subjects. *J. Pharmacol. Exp. Ther.* 358, 125–137.

31. Wolstenholme, A.J., and Rogers, A.T. (2005). Glutamate-gated chloride channels and the mode of action of the avermectin/milbemycinanthelmintics. *Parasitology* 131(Suppl), S85–S95.
32. Yu, Y., Logovinsky, V., Schuck, E., Kaplow, J., Chang, M.K., Miyagawa, T., Wong, N., and Ferry, J. (2014). Safety, tolerability, pharmacokinetics, and pharmacodynamics of the novel gammasecretasemodulator, E2212, in healthy human subjects. *J. Clin. Pharmacol.* 54, 528–536.